Evaluating the anticancer effects of high-dilution preparations of carcinogens such as HIV virus, Hepatitis C virus, Ethanol and Cancer tissues in in-vitro models

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Abstract

Cancer is one of the leading causes of mortality. The recent experiments with high-diluted preparations have shown anticancer effects in in-vitro and vivo models. The fundamental principle of homeopathy suggests that the substances capable of producing certain diseases may have a capacity to alter the same disease if used in the ultra-dilute-potentized form. This hypothesis led certain carcinogens for examining their potential anti-cancer efficacy. Sulforhodamine B assay is useful in determining the cytotoxicity in cell-based studies in evaluating anticancer agents. The protocol involved preparation of homeopathy dilutions, incubation of cells with homeopathy dilutions, SRB binding, and measurement of absorbance. Cells were treated with 30 potencies of HIV nosode, Hepatitis C nosode, Carcinosin, Cancer nosode, and Ethanol along with positive control (Adriamycin). The preparations were tested in HeLa, HepG2, A549, MCF 7, T 24, Jurkat, SCC 40, and HL-60 cell-lines. The homeopathic preparations have shown the anticancer activity measured as percentage growth inhibition. All the homeopathy preparations studied, exhibited anticancer activity on HeLa, HepG2, A 549, T 24, and HL-60 cells. Carcinosin showed the anticancer activity on the SCC 40 cells. Hepatitis C nosode, Carcinosin, and Cancer nosode have shown the anticancer activity on breast cancer cell line MCF-7. None of the preparations exhibited anticancer activity on Human Leukemia Cell Line. High-dilution, potentized preparations of certain carcinogens have demonstrated anti-cancer, cytotoxic effects in the cell-line model, supporting the rationale of the fundamental homeopathic principle the Law of Similars, opening windows to its wider applications in healthcare.

Keywords: Cancer, Homeopathy, HIV nosode, Hepatitis C nosode, Cancer nosode, Ethanol, Law of similar

Introduction

Cancer is a difficult disease yet not fully treatable, calling for newer therapeutic options. Some recent studies with the high-dilution homeopathic drugs have demonstrated anticancer effects in cell line models[1,2,3,4]. Cancer regression in animal models[5] has been demonstrated using high-dilution preparations. More research is encouraged in this direction.

The system of homeopathy was introduced by Samuel Hahnemann, incidentally in the same year 1796 when Edward Jenner [6] introduced vaccinations. Both the approaches are based broadly on the observation that the substances (or organisms) capable of producing diseases in humans, also possibly have the ability to alter the same disease, if administered in an attenuated form (in case of
vaccines) or potentized form in case of homeopathy. In homeopathy, this observation is collated as the *Law of Similars*. Towards substantiating the basic homeopathic principle, certain known carcinogens have been selected for examining their potential anticancer efficacy.

The nosodes are broad-spectrum, widely used homeopathic preparations sourced from biological materials such as cultures or clinical samples of microorganisms (e.g. bacteria, fungi and viruses) or from parasites, diseased tissues (cancerous tissues), or decomposition products from humans or animals [7]. Some nosodes have exhibited anti-disease activity against the diseases they were sourced from. The nosodes prepared from the HIV virus [8], Hepatitis C virus [9] and Plasmodium falciparum parasites [10] have shown respective anti-disease activities in clinical-trials or cell-line models, validating the *Law of Similars*.

HIV infection associated malignancies are common. The HIV virus depresses the immune system and may potentially induce cancers such as Kaposi sarcoma, Non-Hodgkin lymphoma, and cervical cancer [11]. The Hepatitis C virus-infected individuals are at higher risk of developing liver cancer [12]. Ethyl alcohol (C₂H₅O)(subsequently called as Ethanol) has been classified as Group I carcinogen by the International Agency for Research on Cancer (IARC)[13] which can induce female breast, colorectum, larynx, liver, esophagus, oral cavity, and pharynx cancers [14]. As per the principle of the "Law of Similars", the HIV virus, the Hepatitis C virus, and Potentized Ethanol may have a potential to work against malignancies, if administered in the high-dilution, potentized dose. Our earlier studies with Khuda-Bukhsh et al. have corroborated this principle by demonstrating anticancer effects of the HIV nosode [15] and Hepatitis C nosode [16] in cell-line models. We decided to repeat the part of the study and also add three more medicinal substances (Potentized Ethanol and new Cancer Nosode) as well as one of the popular, old homeopathic medicines prepared from undocumented cancer tissues (called Carcinosin), for examining their potential anticancer effects.

In-vitro cell-based cytotoxicity assay promises to be a useful, reliable and rapid method for demonstrating chemotherapeutic drug activity [17]. The MTT/MTS [18] and sulforhodamine B (SRB) assays are widely used techniques for measuring drug-induced cytotoxicity and cell proliferation for large-scale drug-screening applications [19]. The SRB assay was developed by Skehan and colleagues is based on the ability of the SRB dye to bind electrostatically and in a pH-dependent manner on protein basic amino-acid residues. Under mild acidic conditions, SRB binds to protein basic amino-acid residues of trichloroacetic acid (TCA)-fixed cells. It can be quantitatively extracted from cells and solubilized for optical density (OD) measurement by weak bases such as Tris base. Results of the SRB assay were linear with the number of cells and the cellular protein measured at cellular densities in 96-well microtiter plates ranging from 1 to 200% of confluence [20]. The SRB method is appropriate, sensitive and the end point is colorimetric, nondestructive, and indefinitely stable. This method is used for cell density determination, based on the measurement of cellular protein content. The method described here has been optimized for the toxicity screening of compounds to adherent cells in a 96-well format [21].

**Materials and methods**

**Procuring the medicinal substances**

HIV nosode (prepared using sera of HIV type I and type II virus, obtained from HIV infected patients, protocol number LF/001/09) [7], Hepatitis C nosode (prepared using sera of HCV type I and type III virus, obtained from HCV infected patients, protocol number LF/002/09) [8], and Cancer nosode has been developed by the author in the recent years by following the HPI (Homoeopathic Pharmacopeia of India) and other guidelines [22]. Cancer nosode was prepared by the author from several cancer
tissues, including Squamous cell carcinoma (skin), Adenocarcinoma (rectum), and more. Signed Informed consents (approved by ethics committee) of donors for use of biological materials (sera and tissues) were obtained and documented. Carcinosin is a known homeopathic nosode procured from the market, which does not have adequate source information. (SBL, batch no. H1413685). Ethanol (92% dispensing alcohol as per HPI standards) was prepared up to the 30c potency by the author. The use of nosodes for the in-vitro SRB assay was approved by ethics committee, protocol number ACTREC/LF/14/15).

**Homeopathic preparations**

The cell lines were treated with the 30c potency of the following high-dilution preparations:

- a. HIV nosode (HNOD07) prepared from type I and type II HIV infected individual’s serum as per the Homeopathy Pharmacopoeia of India method (7).
- b. Hepatitis C nosode (HCNOD8) prepared from type I and type III Hepatitis C infected individual’s serum as per the Homeopathy Pharmacopoeia of India method (8).
- c. Carcinosin (CSIN01) purchased from Sharda Boiron Laboratories Ltd. (SBL).
- d. Cancer nosode (CNOD02) prepared from cancer tissues as per the Homeopathy Pharmacopoeia of India method.
- e. Ethanol (APOT03) purchased from Merck, Germany, which was potentized.
- f. Adriamycin/ADR (Doxorubicin as a positive control).
- g. Cell control.

The all above preparations were diluted using alcohol as a vehicle which is purchased from Merck (purity of 99.9%) in aseptic conditions and potentized using standardized electro-mechanical potentizer.

**Cell Lines**

Human cervical cancer cell line (HeLa), Hepatoma cell line (Hep-G2), Lung cell carcinoma (A-549), Breast cancer cell line (MCF-7), Bladder cancer cell line (T-24), Leukemia cell line (Jurkat), Oral squamous cell line (SCC-40), and Myeloid Leukemia cell line (HL-60) were used in the study.

**Selection of dose**

Cell lines were treated with the 30c potency of HIV nosode, Hepatitis C nosode, Carcinosin, Cancer nosode, and Potentized Ethanol using the dilutions 1:10, 1:05, 1:01 and direct (30c).

**SRB Assay**

The SRB assay technique as per the development therapeutic program by National Cancer Institute was used as a reference. The Advanced Centre for Treatment, Research and Education in Cancer (ACTREC) provides commercial service for screening anticancer drugs. The work was done through the customized service at ACTREC.

The cell lines were grown in RPMI 1640 (Roswell Park Memorial Institute) medium containing 10% fetal bovine serum and 2 mM L-glutamine. For present screening experiments, the cells were
cultivated into 96 well microtiter plates in 100 μL at plating densities of cell count +5000. The initial cell seeding density was selected depending on the doubling time of the individual cell lines (23). Plates were incubated at 37°C, 5 % CO2, 95 % air and 100 % relative humidity for 24 hours prior to addition of experimental drugs.

The homeopathic samples (30c potency; direct) were stored at room temperature, diluted to 1:10, 1:05 and 1:01 with vehicle (un-potentized alcohol). Aliquots of 10 μl of these different homeopathy dilutions were added to the appropriate well already containing 90 μl of the medium, resulting in the required final drug concentrations i.e. 10 μg/ml, 20 μg/ml, 40 μg/ml, 80 μg/ml.

After compound addition, plates were incubated at standard conditions for 48 hours and the assay was terminated by the addition of cold Trichloroacetic Acid (TCA). Cells were fixed in-situ by the gentle addition of 50 μl of cold 30 % (w/v) TCA (final concentration, 10 % TCA) and incubated for 60 minutes at 4°C. The supernatant was discarded; the plates were washed five times with water and air dried. The SRB solution (50 μl) at 0.4 % (w/v) in 1 % acetic acid was added to each of the wells, and plates were incubated for 20 minutes at room temperature. After staining, the unbound dye was recovered and the residual dye was removed by washing five times with 1 % acetic acid. The plates were air dried. The bound stain was subsequently eluted with 10 mM trizma base, and the absorbance was read on a plate reader at a wavelength of 540 nm with 690 nm reference wavelength.

Percent growth was calculated on a plate-by-plate basis for test wells relative to control wells. Percent growth was expressed as the ratio of average absorbance of the test well to the average absorbance of the control wells X 100.

Using the six absorbance measurements [time zero (Tz), control growth (C), and test growth in the presence of drug at the four concentration levels (Ti)], the percentage growth was calculated at each of the drug concentration levels. Percentage growth inhibition was calculated as [Ti/C] x 100 %.

Growth inhibition of 50 % (GI50) is calculated from \([\frac{(Ti-Tz)}{(C-Tz)}] \times 100 = 50\), which is the drug concentration resulting in a 50% reduction in the net protein increase (as measured by SRB staining) in control cells during the drug incubation. The drug concentration resulting in total growth inhibition (TGI) is calculated from \(Ti = Tz\).

The experiment was conducted in triplicate for each cell line to validate the results.

Results

Images 1-3 showing cell growth after SRB staining in A-549, MCF-7 and T-24 cell line indicating efficacy of test samples compared with controls.
The figures 1 to 9 represent the percentage control growth studied in all cell lines for HIV nosode, Hepatitis C, Carcinosin and Cancer nosode, Potentized Ethanol in 30c dilution, as 1:10, 1:05, 1:01 and direct (30c).

Figure 1 % Control Growth and Drug Concentrations (HeLa)
**Figure 2** % Control Growth and Drug Concentrations (Hep-G2)

**Figure 3** % Control Growth and Drug Concentrations (A-549)

**Figure 4** % Control Growth and Drug Concentrations (MCF-7)
Figure 5 % Control Growth and Drug Concentrations (T-24)

Figure 6 % Control Growth and Drug Concentrations (JURKAT)

Figure 7 % Control Growth and Drug Concentrations (SCC-40)
All the five homeopathy preparations have shown the anticancer activity on Human cervical cancer cell line HeLa. (figures 10, 1 and 9).

Figure 10 Growth curve- Human Cervical cancer cell line (HeLa)
Human Hepatoma Cell Line Hep-G2

All the five homeopathy preparations have shown the anticancer activity on human hepatoma cell line (Hep-G2). (Figures 11, 2 and 9).

**Figure 11** Growth curve- Human Hepatoma cell line (Hep-G2)

![Growth Curve: Human Hepatoma Cell Line Hep-G2](image)

Human Lung Cell Carcinoma Cell A-549:

All the five homeopathy preparations have shown the anticancer activity on human lung cell carcinoma cell (A-549) (Refer figure 12, 3 and 9).

**Figure 12** Growth curve-Human lung cell carcinoma cell (A-549)

![Growth Curve: Human Lung Cell Carcinoma Cell A-549](image)

Human Breast Cancer Cell Line MCF-7

Hepatitis C, Carcinosin, and Cancer nosode have shown the anticancer activity on breast cancer cell line MCF-7 (Figures 13, 4 and 9).

**Figure 13** Growth curve- Human breast cancer cell line MCF-7

![Growth Curve: Human Breast Cancer Cell Line MCF-7](image)
Human Bladder Cancer Cell Line T-24

All the five homeopathy preparations have shown the anticancer activity on human bladder cancer cell line (T-24) (Figures 14, 5 and 9).

Figure 14 Growth curve-human bladder cancer cell line (T-24)

Human Leukemia Cell Line JURKAT

None of the preparations has shown any anticancer activity in the Human Leukemia Cell Line (Figures 15, 6 and 9).

Figure 15 growth curve-Human Leukemia Cell Line JURKAT

Human Oral Squamous Cell Line (SCC-40)

Only Carcinosin has shown the anticancer activity in the oral squamous cell line (Figures 16, 7 and 9).

Figure 16 Growth curve- Human oral squamous cell line (SCC-40)
Human Myeloid Leukemia Cell Line HL-60

All the five preparations have shown the anticancer activity on the Human Myeloid Leukemia cell line. (Figures 17, 8 and 9).

Figure 17 growth curve- Human Myeloid Leukemia cell line (HL-60)

The high-dilutions can be prepared by the process of potentization using either alcohol or water (Water for injection- WFI) as a medium. Another set of high dilution preparations of all the test drugs was prepared separately using water (WFI) as a vehicle for undertaking a study to see the difference in results if any. The preparations thus made were tested on Human Lung Cancer cell line (A-549); only one cell line was used to check if the nature of vehicle has any impact on the activity. No anticancer activity was observed in any of the test drugs prepared using the WFI as a vehicle. Similarly, to determine if it is the effect of alcohol or that of the process of potentization that produced anticancer effect, another study was conducted to examine the effect of Ethanol (without potentization) on Human Lung Cancer cell line (A-549). The un-potentized alcohol did not show any anticancer activity on A-549. (figure 18 and table 1)

Figure 18: Growth curve of A-549
Table 1: Results for water-based preparations and un-potentized alcohol on Human Lung Cancer cell line (A-549)

<table>
<thead>
<tr>
<th>Medicine</th>
<th>% Control Growth and Drug Concentrations (A-549)</th>
<th>Drug concentrations (%v/v) calculated from graph</th>
</tr>
</thead>
<tbody>
<tr>
<td>% v/v</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>HIV Nosode</td>
<td>107.1</td>
<td>105.1</td>
</tr>
<tr>
<td>Hep C nosode</td>
<td>108.5</td>
<td>106.1</td>
</tr>
<tr>
<td>Carcinosin</td>
<td>107.5</td>
<td>104.4</td>
</tr>
<tr>
<td>Cancer nosode</td>
<td>112.0</td>
<td>107.6</td>
</tr>
<tr>
<td>Un-potentized alcohol</td>
<td>111.4</td>
<td>110.9</td>
</tr>
<tr>
<td>Adriamycin</td>
<td>-5.0</td>
<td>-7.4</td>
</tr>
</tbody>
</table>

Discussion

The research evaluating the efficacy of the high-dilution and potentized medicines, that too in the lab models outside of the human system, is relatively novel. The controlled study has shown three homeopathic drugs sourced from carcinogens such as HIV virus, Hepatitis C virus, and Ethanol; and two from cancer tissues (new Cancer nosode, and Carcinosin) in high dilutions (Potentized) having variable anticancer activities in Human cervical cancer (HeLa), Hepatoma (HepG2), Lung cell carcinoma A 549, Bladder cancer T 24 and Myeloid Leukemia HL-60.

On investigative communication by the author with some of the leading manufactures of Carcinosin nosodes, it was found that the currently available nosode is often a mix of over 28 different cancer tissues, of which no source details are available with the manufactures [24]. The author has prepared
a new Cancer Nosode from well-defined tissues using five different cancers (The article is under review for publication.)

The New Cancer nosode has shown anticancer effects on HeLa, Hep-G2, A-549, MCF-7, T-24, and HL-60 cell lines while Carcinosin has shown effects against HeLa, Hep-G2, A-549, MCF-7, T-24, SCC-40 and HL-60 which may be due to a wider spectrum of cancer tissues. More studies with individual tissues as well as multiple tissues may prove more insightful.

Only Carcinosin has shown anticancer activity in the oral squamous cell line (SCC-40). None of the homeopathic preparations has shown any anticancer activity in Human Leukemia Cell Line (Jurkat). Hepatitis C nosode, Carcinosin, and Cancer nosode have shown the anticancer activity against breast cancer cell line MCF-7.

Potentized Ethanol has shown anticancer activity in HeLa, Hep-G2, A-549, T-24 and HL-60 cell lines. An additional study using the un-potentized Ethanol did not show any anticancer effect in A-549 cell line, suggesting the role of potentization in rendering the therapeutic effect of un-potentized alcohol, while the Potentized Ethanol did show anticancer activity on the same cell line.

Potentized Ethanol has not shown any activity in MCF-7 and SCC 40 cell line, whereas Hepatitis C nosode, Cancer nosode (both were Potentized using the same ethanol as a vehicle) and Carcinosin have shown activity in MCF-7 and Carcinosin in SCC 40. This suggests that the positive effect in all the three nosodes was not due to the Potentized Ethanol but due to the respective Potentized source materials. This finding was supported by the outcome of the study done in past where HIV nosode and Hepatitis C nosode have shown anticancer activity on MCF-7 and A549 cell lines as compared to Potentized Ethanol [14,15].

The same preparations when made using water (Water for injection) as a medium instead of alcohol, did not show any anticancer activity. This observation suggests that alcohol as a medium seems to retain the therapeutic properties of the original drug substance better than only water. Literature suggests the use of ethanol-water for serial dilutions. Many theories have put up to describe the role of water-ethanol clusters, transfer of information of specific remedy material in water-ethanol colloidal solution [25].

HIV [26], Hepatitis C virus [27] and Potentized Ethanol [28,29] are known carcinogens. The Cancer tissues are end products of the malignant process, which may include a wide range of proteins, debris, cytokines, immunocytes, cells, and more. The ultra-diluted preparations of all of these carcinogens and cancer tissues have shown anticancer effects. More carcinogens and cancer tissues using more cell line models as well as in-vivo studies may lead to understanding the mode of actions.

**Conclusion**

High dilution, potentized preparations of certain carcinogens have demonstrated anti-cancer, cytotoxic effects in the cell-line model, supporting the rationale of the fundamental homeopathic principle the *Law of Similars*, opening windows to its wider applications in healthcare.

**Acknowledgments**

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References


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